

# CERTIFICATION

## **AOAC<sup>®</sup>** *Performance Tested*<sup>SM</sup>

Certificate No. 061502

The AOAC Research Institute hereby certifies the test kit known as:

### **GlutenTox®** Pro

manufactured by

Hygiena Diagnóstica España P. I. Parque Plata, Calle Cañada Real 31-35 Camas, Sevilla 41900 Spain

This method has been evaluated in the AOAC<sup>®</sup> *Performance Tested Methods*<sup>SM</sup> Program and found to perform as stated by the manufacturer contingent to the comments contained in the manuscript. This certificate means that an AOAC<sup>®</sup> Certification Mark License Agreement has been executed which authorizes the manufacturer to display the AOAC *Performance Tested* <sup>SM</sup> certification mark along with the statement - "THIS METHOD'S PERFORMANCE WAS REVIEWED BY AOAC RESEARCH INSTITUTE AND WAS FOUND TO PERFORM TO THE MANUFACTURER'S SPECIFICATIONS" - on the above-mentioned method for a period of one calendar year from the date of this certificate (October 13, 2021 – December 31, 2022). Renewal may be granted at the end of one year under the rules stated in the licensing agreement.

Scott Coates

Scott Coates, Senior Director Signature for AOAC Research Institute October 13, 2021 Date

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METHOD AUTHORS Miguel A. Síglez, Bárbara Nocea, María del Mar Pérez, Eva Mª García, Laura León, Carlos Galera MODIFCATION DECEMBER 2018: Hygiena Diagnóstica España	Biomedal, S. L. Avenida Américo Vespucio, 5-E, 1ª M-12 41092 Sevilla Spain	CURRENT COMPANY Hygiena Diagnóstica España P. I. Parque Plata, Calle Cañada Real 31-35 Camas, Sevilla 41900 Spain
KIT NAME(S) GlutenTox® Pro	<b>CATALOG NUMBERS</b> KIT 3000 (KT-5660; 25 analysis); KIT 3001 (KT-528)	3; 5 analysis)
INDEPENDENT LABORATORY Q Laboratories, Inc. 1400 Harrison Ave Cincinnati, OH 45214 USA	AOAC EXPERTS AND PEER REVIEWERS Joe Boison <sup>1</sup> , Mary Trucksess <sup>2</sup> , Terry Koener <sup>3</sup> <sup>1</sup> Canadian Food Inspection Agency, Saskatooan, C <sup>2</sup> Mycotoxin Consultant, Virginia, USA <sup>3</sup> Health Canada	anada
APPLICABILITY OF METHOD Target analyte – Gluten Matrixes – bread, rice flour, paté, rolled oat, yogurt, food-grade painted wood, plastic, rubber, sealed ceramic, stainless steel Performance claims - The GlutenTox®Pro test kit is a quick and easy to use screening method for the detection of gluten in raw or cooked foods and on environmental surfaces. The method is specific and reliable and provides sensitive and accurate test results comparable to AOAC OMA 2012.01.	REFERENCE METHOD AOAC Official Methods of Analysis (OMA) 2012.0 Gluten in Foods Containing Wheat, Rye, and Barl	
ORIGINAL CERTIFICATION DATE June 26, 2015	CERTIFICATION RENEWAL RECORD Renewed annually through December 2022	
METHOD MODIFICATION RECORD 1. December 2018 Level 2 2. November 2019 Level 1	<ul> <li>SUMMARY OF MODIFICATION</li> <li>1. Purchase and location change from Bi Vespucio, 5-E, 1<sup>a</sup> M-12, 41092 Sevilla, Diagnóstica España P. I. Parque Plata, 41900 Camas, Sevilla, Spain.</li> <li>2. Editorial/clerical changes.</li> </ul>	Spain to Hygiena
Under this AOAC <sup>®</sup> <i>Performance Tested</i> <sup>5M</sup> License Number, 061502 this method is distributed by: NONE	Under this AOAC <sup>®</sup> <i>Performance Tested<sup>sm</sup></i> License method is distributed as: NONE	Number, 061502 this

#### PRINCIPLE OF THE METHOD (1)

The GlutenTox<sup>®</sup>Pro method is an immunochromatographic assay for the detection of gluten in food and beverages (with non-hydrolyzed gluten) with different composition and levels of processing, from raw materials to processed food. In addition, the GlutenTox<sup>®</sup>Pro Test Kit can be used to control the cleanliness of food production zones through surface analysis, a prerequisite to prevent the risk of cross-contamination in the final product.

#### **DISCUSSION OF THE VALIDATION STUDY (1)**

The GlutenTox<sup>®</sup>Pro method did not show cross-reactivity to any of the compounds included in the list of *Validation Procedures for Quantitative Gluten ELISA Methods: AOAC Allergen Community Guidance and Best Practices*<sup>4</sup> used in the production of gluten-free products. The GlutenTox<sup>®</sup>Pro assay also did not show any interference, when tested with the compounds from the list in the presence of gluten. No unexpected results were obtained however gum-type samples can be difficult to analyze due to the thick paste formed when added to the extraction solution provided in the GlutenTox<sup>®</sup>Pro test kit. A warning to this type of samples has been included in the instructions for use.

The GlutenTox<sup>®</sup>Pro test kit performed as expected in the selected food matrixes (rice flour, bread, rolled oat, pâté and yogurt) and test conditions (spike level and detection threshold combinations), 5 ppm being the lowest concentration of gluten that can be detected with the kit.

In all matrixes tested, the GlutenTox<sup>®</sup>Pro method demonstrated 100 % specificity [probability of detection (POD) 0.00. confidence interval (CI) 0.00-0.11] at 0 ppm spiked level of gluten and 100 % sensitivity (POD 1.00., CI 0.89-1.00) at each spiked level of gluten and threshold level combinations. No false negative results were obtained in the food matrix study. The assay did not experience hook effect at any threshold level tested when the rice flour matrix was spiked at very high spiked levels of gluten (10,000 ppm).

In the incurred sample study, the incurred residue target level was approximately 25 ppm of gluten, the initial spiking level in the uncooked matrix was 50 ppm of gluten and a 78.2 % recovery was obtained when tested with the AOAC OMA 2012.01 method<sup>11</sup> (recovery could be between 50-150%).

The GlutenTox<sup>®</sup>Pro test kit performed as expected in the incurred bread sample and the results obtained in the incurred matrix study were consistent with those obtained in the selected food matrix study with bread. In both studies, false negative and/or overestimated results were not observed.

The results obtained when the GlutenTox®Pro test kit was tested with the selected environmental surfaces (food-grade painted wood, plastic, rubber, sealed ceramic and stainless steel) demonstrated a 100 % specificity (POD 0.00, Cl0.00-0.11) at the unspiked level of gluten contamination and a 100 % sensitivity (POD 1.00., Cl 0.89-1.00) at the high level of gluten contamination (400 ng/16 cm<sup>2</sup>), in each of the environmental surfaces analyzed.

At the low level of gluten contamination (16 ng/16 cm<sup>2</sup>), the GlutenTox<sup>®</sup>Pro assay was able to detect as little as 16 ng of gluten when analyzed with the environmental surface matrixes.

The lot-to-lot data, the accelerated stability data (10 days, 20 days, 35 days, 50 days and 90 days at 42°C) and the real time stability data (3 months, 18 months, 24 months, and 30 months at 22°C) showed evidence that the GlutenTox®Pro method is stable and can be consistently manufactured with reproducible quality.

Test kit variation data among 3 test kits of a single lot of GlutenTox<sup>®</sup>Pro test kit demonstrated no statistical difference in gluten detection between the test kits. Occasional slight overestimations are irrelevant in gluten analysis compared to a problem that could arise from false negatives or underestimations. No false negative results were observed in the entire validation study.

Robustness data indicated that the GlutenTox<sup>®</sup>Pro assay remained unaffected by minor variations in procedural parameters with the exception of the amount of time that the test strip was left in the dilution sample solution before reading the result. Due to the test format, there must be sufficient time for the dilution sample solution to travel up the test strip, and this time cannot be shortened. The effect of decreasing the strip incubation time was not dependent of the amount of dilution sample solution used but this effect was smaller when coupled with an increased sample extraction time. When the test strip was left in a smaller amount of dilution sample solution some invalid results appeared.

30

30

0

1.00

1.00

0.00

0.89, 1.00

0.89, 1.00

39.1

Variance (σ²)

1.2

Madulu	Gluten Spiked	Detection Threshold	Na		Candio	late	Ave. AOAC OMA
Matrix	Level*	(ppm)	IN <sup>o</sup>	Xp	PODc <sup>c</sup>	95% Cl <sup>d</sup>	2012.01 results, ppm gluten, N=3
		5	30	0	0.00	0.00, 0.11	
	0	10	30	0	0.00	0.00, 0.11	
	0 ppm	20	30	0	0.00	0.00, 0.11	<2.5
Incurred Matrix		40	30	0	0.00	0.00, 0.11	1
(Bread)		5	30	30	1.00	0.89, 1.00	
(breau)							

30

30

30

#### Table 3: GlutenTox<sup>®</sup>Pro Test Kit Incurred Matrix (Bread) – POD Results (1)

\*Gluten Spiked Level results after cooking the bread

<sup>a</sup>N = Number of test portions

39.1 ppm

<sup>b</sup>x = Number of positive test portions

<sup>c</sup>POD<sub>c</sub> = Candidate method confirmed positive outcomes divided by the total number of trials

10

20

40

<sup>d</sup>95% Confidence Intervals

#### Table 4: GlutenTox<sup>®</sup>Pro Test Kit for Rice Flour – POD Results (1)

	Gluten Spiked	GlutenTox <sup>®</sup> Pro Detection	Na		Candida	te	Ave. AOAC OMA	Variance
Matrix	Level	Threshold (ppm gluten)	N°	xb	PODc <sup>c</sup>	95% Cl <sup>d</sup>	2012.01 results, ppm gluten, N=3	(σ²)
		5	30	0	0.00	0.00, 0.11		
	0.555	10	30	0	0.00	0.00, 0.11		-
	0 ppm	20	30	0	0.00	0.00, 0.11	<2.5	
		40	30	0	0.00	0.00, 0.11		
		5	30	11	0.37	0.22, 0.54		
Rice Flour	2 nnm	10	30	0	0.00	0.00, 0.11		
	3 ppm	20	30	0	0.00	0.00, 0.11	3.9	0.2
		40	30	0	0.00	0.00, 0.11		
		5	30	30	1.00	0.89, 1.00		
	8 ppm	10	30	3	0.10	0.03, 0.26		
		20	30	0	0.00	0.00, 0.11	8.8	0.2

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		40	30	0	0.00	0.00, 0.11		
		5	30	30	1.00	0.89, 1.00		
	15	10	30	30	1.00	0.89, 1.00		
	15 ppm	20	30	0	0.00	0.00, 0.11	14.5	0.3
		40	30	0	0.00	0.00, 0.11		
		5	30	30	1.00	0.89, 1.00		
	25	10	30	30	1.00	0.89, 1.00		
	25 ppm	20	30	30	1.00	0.89, 1.00	21.5	1.8
		40	30	0	0.00	0.00, 0.11		
		5	30	30	1.00	0.89, 1.00		
	45	10	30	30	1.00	0.89, 1.00		
	45 ppm	20	30	30	1.00	0.89, 1.00	38.0	1.1
		40	30	30	1.00	0.89, 1.00		
		5	10	10	1.00	0.72, 1.00		
	10.000	10	10	10	1.00	0.72, 1.00		
	10,000 ppm	20	10	10	1.00	0.72, 1.00	8061.0	-
		40	10	10	1.00	0.72, 1.00		
201	Numera a strategy of							

<sup>a</sup>N = Number of test portions

<sup>b</sup>x = Number of positive test portions

 $^{c}\text{POD}_{c}$  = Candidate method confirmed positive outcomes divided by the total number of trials

<sup>d</sup>95% Confidence Intervals

Matrix	Gluten Spiked	GlutenTox <sup>®</sup> Pro Detection	Nª		Candio	date	Ave. AOAC OMA 2012.01 results, ppm	Variance
Level	Level	Threshold (ppm gluten)	IN	Xp	PODc <sup>c</sup>	95% Cl <sup>d</sup>	gluten, N=3	(σ²)
		5	30	0	0.00	0.00, 0.11		
	0	10	30	0	0.00	0.00, 0.11		_
	0 ppm	20	30	0	0.00	0.00, 0.11	<2.5	
		40	30	0	0.00	0.00, 0.11		
		5	30	0	0.00	0.00, 0.11		
	3 ppm	10	30	0	0.00	0.00, 0.11		
	3 ppm	20	30	0	0.00	0.00, 0.11	2.3	0.1
		40	30	0	0.00	0.00, 0.11		
		5	30	30	1.00	0.89, 1.00		
		10	30	0	0.00	0.00, 0.11		
	8 ppm	20	30	0	0.00	0.00, 0.11	7.2	0.1
Bread		40	30	0	0.00	0.00, 0.11		
Diedu		5	30	30	1.00	0.89, 1.00		
	15 ppm	10	30	30	1.00	0.89, 1.00		
	13 hhiu	20	30	0	0.00	0.00, 0.11	14.0	1.5
		40	30	0	0.00	0.00, 0.11		
		5	30	30	1.00	0.89, 1.00		
	25	10	30	30	1.00	0.89, 1.00		
	25 ppm	20	30	30	1.00	0.89, 1.00	21.1	2.5
		40	30	0	0.00	0.00, 0.11		
		5	30	30	1.00	0.89, 1.00		
	45 mm	10	30	30	1.00	0.89, 1.00	7	
	45 ppm	20	30	30	1.00	0.89, 1.00	38.5	2.4
		40	30	30	1.00	0.89, 1.00		

<sup>a</sup>N = Number of test portions

<sup>b</sup>x = Number of positive test portions

<sup>c</sup>POD<sub>c</sub> = Candidate method confirmed positive outcomes divided by the total number of trials

<sup>d</sup>95% Confidence Intervals

Matrix	Gluten Spiked	GlutenTox <sup>®</sup> Pro Detection	Nª		Candio	date	Ave. AOAC OMA	Varianc
Watrix	Level	Threshold (ppm gluten)	N-	Xp	PODc <sup>c</sup>	95% Cl <sup>d</sup>	2012.01 results, ppm gluten, N=3	(σ²)
		5	30	0	0.00	0.00, 0.11		
	0 ppm	10	30	0	0.00	0.00, 0.11		_
	0 ppm	20	30	0	0.00	0.00, 0.11	<2.5	
		40	30	0	0.00	0.00, 0.11		
		5	30	0	0.00	0.00, 0.11		
	2	10	30	2	0.07	0.02, 0.21		
	3 ppm	20	30	0	0.00	0.00, 0.11	2.7	0.0
		40	30	0	0.00	0.00, 0.11		
		5	30	30	1.00	0.89, 1.00		
		10	30	0	0.00	0.00, 0.11		
	8 ppm	20	30	0	0.00	0.00, 0.11	8.3	1.7
Rolled oat		40	30	0	0.00	0.00, 0.11		
Noneu oat		5	30	30	1.00	0.89, 1.00		
	15 0000	10	30	30	1.00	0.89, 1.00		
	15 ppm	20	30	0	0.00	0.00, 0.11	12.6	1.0
		40	30	0	0.00	0.00, 0.11		
		5	30	30	1.00	0.89, 1.00		
	25	10	30	30	1.00	0.89, 1.00		
	25 ppm	20	30	30	1.00	0.89, 1.00	20.4	3.4
		40	30	0	0.00	0.00, 0.11		
		5	30	30	1.00	0.89, 1.00		
	45	10	30	30	1.00	0.89, 1.00	1	
	45 ppm	20	30	30	1.00	0.89, 1.00	41.0	3.5
		40	30	30	1.00	0.89, 1.00		

<sup>a</sup>N = Number of test portions

<sup>b</sup>x = Number of positive test portions

 $^{c}\mbox{POD}_{\mbox{C}}$  = Candidate method confirmed positive outcomes divided by the total number of trials d95% Confidence Intervals

Matrix	Gluten Spiked	GlutenTox <sup>®</sup> Pro Detection	Nª		Candio	date	Ave. AOAC OMA 2012.01 results, ppm	Variance
IVIALITA	Level	Threshold (ppm gluten)	N	Xp	PODc <sup>c</sup>	95% Cl <sup>d</sup>	gluten, N=3	(σ²)
		5	30	0	0.00	0.00, 0.11		
	0 ppm	10	30	0	0.00	0.00, 0.11	1	_
	0 ppm	20	30	0	0.00	0.00, 0.11	<2.5	
		40	30	0	0.00	0.00, 0.11		
		5	30	0	0.00	0.00, 0.11		
	3 ppm	10	30	9	0.30	0.17, 0.48	7	
		20	30	0	0.00	0.00, 0.11	3.0	0.7
		40	30	0	0.00	0.00, 0.11		
		5	30	30	1.00	0.89, 1.00		
	maga 8	10	30	0	0.00	0.00, 0.11	]	
	8 ppm	20	30	0	0.00	0.00, 0.11	9.2	0.4
Pâté		40	30	0	0.00	0.00, 0.11		
Pale		5	30	30	1.00	0.89, 1.00		
	15 ppm	10	30	30	1.00	0.89, 1.00		
	12 hhiu	20	30	0	0.00	0.00, 0.11	16.1	0.4
		40	30	0	0.00	0.00, 0.11		
		5	30	30	1.00	0.89, 1.00		
	25	10	30	30	1.00	0.89, 1.00	1	
	25 ppm	20	30	30	1.00	0.89, 1.00	27.6	36.8
		40	30	0	0.00	0.00, 0.11		
		5	30	30	1.00	0.89, 1.00		
	45 nnm	10	30	30	1.00	0.89, 1.00		
	45 ppm	20	30	30	1.00	0.89, 1.00	41.0	18.9
		40	30	30	1.00	0.89, 1.00		

<sup>a</sup>N = Number of test portions

<sup>b</sup>x = Number of positive test portions

<sup>c</sup>POD<sub>c</sub> = Candidate method confirmed positive outcomes divided by the total number of trials <sup>d</sup>95% Confidence Intervals

Matrix	Gluten Spiked	GlutenTox <sup>®</sup> Pro Detection	Nª		Candi	date	Ave. AOAC OMA 2012.01 results, ppm	Variance (σ²)
Level	Level	Threshold (ppm gluten)	IN	Xp	PODc <sup>c</sup>	95% Cl <sup>d</sup>	gluten, N=3	(0)
		5	30	0	0.00	0.00, 0.11		
	0	10	30	0	0.00	0.00, 0.11		_
	0 ppm	20	30	0	0.00	0.00, 0.11	<2.5	
		40	30	0	0.00	0.00, 0.11		
		5	30	0	0.00	0.00, 0.11		
	2	10	30	0	0.00	0.00, 0.11		
	3 ppm	20	30	0	0.00	0.00, 0.11	3.2	0.0
		40	30	0	0.00	0.00, 0.11		
		5	30	30	1.00	0.89, 1.00		
		10	30	0	0.00	0.00, 0.11		
	8 ppm	20	30	0	0.00	0.00, 0.11	9.3	0.0
Yogurt		40	30	0	0.00	0.00, 0.11		
roguit		5	30	30	1.00	0.89, 1.00		
	15 ppm	10	30	30	1.00	0.89, 1.00		
	12 bbu	20	30	0	0.00	0.00, 0.11	16.6	2.4
		40	30	0	0.00	0.00, 0.11		
		5	30	30	1.00	0.89, 1.00		
	25	10	30	30	1.00	0.89, 1.00		
	25 ppm	20	30	30	1.00	0.89, 1.00	24.9	0.5
		40	30	0	0.00	0.00, 0.11		
		5	30	30	1.00	0.89, 1.00		
	45	10	30	30	1.00	0.89, 1.00		
	45 ppm	20	30	30	1.00	0.89, 1.00	38.2	1.5
		40	30	30	1.00	0.89, 1.00		

 $^{a}N = Number of test portions$ 

<sup>b</sup>x = Number of positive test portions

 $^{\rm c}\text{POD}_{c}$  = Candidate method confirmed positive outcomes divided by the total number of trials  $^{\rm d}\text{95\%}$  Confidence Intervals

Matrix	Amount of Spiked Gluten	Na		Candidate			
(16 cm²)	(ng/16 cm <sup>2</sup> )	Nª	xb	PODc <sup>c</sup>	95% Cl <sup>d</sup>		
	Blank 0	5	0	0.00	0.00, 0.43		
Food-grade painted wood	Low 16	30	25	0.83	0.66, 0.93		
painteu woou	High 400	5	5	1.00	0.57, 1.00		
	Blank 0	5	0	0.00	0.00, 0.43		
Plastic	Low 16	30	23	0.77	0.59, 0.88		
	High 400	5	5	1.00	0.57, 1.00		
	Blank 0	5	0	0.00	0.00, 0.43		
Rubber	Low 16	30	26	0.87	0.70, 0.95		
	High 400	5	5	1.00	0.57, 1.00		
	Blank 0	5	0	0.00	0.00, 0.43		
Sealed Ceramic	Low 16	30	25	0.83	0.66, 0.93		
	High 400	5	5	1.00	0.57, 1.00		
Stainless steel	Blank 0	5	0	0.00	0.00, 0.43		
	Low 16	30	21	0.70	0.52, 0.83		
	High 400	5	5	1.00	0.57, 1.00		

<sup>a</sup>N = Number of test portions

<sup>b</sup>x = Number of positive test portions

 $^{c}\text{POD}_{c}$  = Candidate method confirmed positive outcomes divided by the total number of trials  $^{d}95\%$  Confidence Intervals

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